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Patentanmeldung Nr. Patent application No. Demande de brevet n°

02021601.6

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Anmeldung Nr:

Application no.: 02021601.6

Demande no:

Anmeldetag:

Date of filing: 27.09.02

Date de dépôt:

Anmelder/Applicant(s)/Demandeur(s):

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SUISSE

Bezeichnung der Erfindung/Title of the invention/Titre de l'invention:

(Falls die Bezeichnung der Erfindung nicht angegeben ist, siehe Beschreibung.

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Process for producing vitamin B6

In Anspruch genommene Priorität(en) / Priority(ies) claimed /Priorité(s)
revendiquée(s)

Staat/Tag/Aktenzeichen/State/Date/File no./Pays/Date/Numéro de dépôt:

Internationale Patentklassifikation/International Patent Classification/
Classification internationale des brevets:

C12P/

Am Anmeldetag benannte Vertragstaaten/Contracting states designated at date of
filing/Etats contractants désignées lors du dépôt:

AT BE BG CH CY CZ DE DK EE ES FI FR GB GR IE IT LI LU MC NL PT SE SK TR

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Case 21413Process for producing vitamin B₆

The present invention relates to a novel microorganism and a process for preparing vitamin B₆ by using the same.

5 "Vitamin B₆" as used in the present invention includes pyridoxol, pyridoxal, and pyridoxamine. Vitamin B₆ is a vitamin indispensable to human beings or other animals and used as a raw material of medicines or as feed additives. As a process for preparing vitamin B₆ in a fermentation method, a process for preparing vitamin B₆ using a microorganism derived from the genus *Sinorhizobium* (also known as *Rhizobium*) (EP 765,938) is known. But it is necessary to construct a new microorganism with a higher yield of vitamin B₆ and
10 to develop an improved industrial fermentation process which can produce vitamin B₆ with sufficiently high production efficiency using said microorganism.

According to the present invention, it is possible to produce vitamin B₆ more efficiently than the previous process. The present inventors first constructed a recombinant microorganism of the genus *Sinorhizobium* capable of producing vitamin B₆ incorporated with a
15 recombinant plasmid comprising a vector containing pyridoxol 5'-phosphate synthase gene (referred to as *pdxJ* hereinafter). This recombinant microorganism showed an increased production of vitamin B₆. This recombinant microorganism was further mutated to acquire a phenotypic property of histidine requirement, or glycine resistance. Such a mutant microorganism shows further increased productivity of vitamin B₆. A mutant
20 which acquired the above mentioned two phenotypic properties simultaneously shows drastically increased productivity of vitamin B₆, and that vitamin B₆ can advantageously be produced in the culture broth by cultivating the microorganism, and can be recovered therefrom in a desired purity.

Hei/fm; 27.09.2002

The present invention provides a mutant of a recombinant microorganism of the genus *Sinorhizobium* capable of producing vitamin B₆ having a recombinant plasmid with pdxJ gene that acquired a phenotypic property of histidine requirement or glycine resistance, or combination of the phenotypic properties thereof.

- 5 Another object of the present invention is to provide a process for preparing vitamin B₆ which comprises cultivating said microorganism in a culture medium and collecting the produced vitamin B₆.

As a parent strain for preparing a microorganism in the present invention, any strains belonging to the genus *Sinorhizobium* capable of producing vitamin B₆ can be used, and a
10 microorganism belonging to the genus *Sinorhizobium* may be isolated from natural sources, or may be purchased from culture collections. *S. meliloti* IFO 14782 (DSM 10226) is preferable to the present invention. A microorganism capable of producing large amount of vitamin B₆ can be constructed as described below.

(1) Preparation of *S. meliloti* IFO 14782 Harboring Recombinant Plasmid with pdxJ Gene

15 [A] Construction of expression plasmid for pdxJ

"pdxJ" referred to herein means the gene encoding an enzyme catalyzing synthesis of pyridoxol 5'-phosphate from 1-deoxy-D-xylulose 5-phosphate and aminoacetone 3-phosphate. A pdxJ-gene derived from microorganism belonging to the genus *Sinorhizobium* is preferable. For example, a DNA of pdxJ derived from *S. meliloti* IFO 14782 can be cloned
20 in the following manner. The primers for polymerase chain reaction (referred to as PCR hereinafter) are synthesized in accordance with the DNA sequence of pdxJ in a DNA database of *S. meliloti* strain 1021, and which contain restriction enzyme recognition site at the 5' end of each primer. The pdxJ gene can be amplified by PCR using the primers and chromosomal DNA of *S. meliloti* IFO 14782. Amplified pdxJ is ligated into a vector
25 replicable in *Escherichia coli* such as available pUC series or pBR series. A plasmid, wherein pdxJ is inserted, can be selected by agarose gel analysis of the plasmid digested with endonuclease, and the sequence of amplified region can be ascertained with a DNA sequencer.

As a vector for expression of PdxJ protein in *E. coli*, a vector can be remodeled into a new
30 plasmid, which has a promoter functioning in *E. coli* such as ptac, ptrp, plac, or ptrc followed by restriction enzyme recognition sequence. A plasmid expressing the PdxJ protein in *E. coli* can be provided by inserting the thus-obtained pdxJ into a thus-obtained expression plasmid, which encodes pdxJ under control of a promoter.

As a vector for expression of PdxJ protein in *S. meliloti*, a broad-host range vector, such as pVK100, pRK290, pLAFR1 or RSF1010, can be used. A plasmid expressing PdxJ protein in *S. meliloti* can be provided by inserting a DNA fragment encoding a promoter functioning in *S. meliloti*, such as ptac, plac, ptrc, pS1 (promoter of small ribosomal subunit of *S. meli-*
5 *loti*), or pNm (promoter of neomycin-resistant gene) and pdxJ into a broad-host range vector.

The procedure for constructing recombinant vectors can be performed according to standard techniques known in the fields of molecular biology, bioengineering, and genetic engineering.

10 [B] Introduction of recombinant plasmid with pdxJ into *S. meliloti* IFO 14782

A plasmid encoding pdxJ of *S. meliloti* can be transformed to *E. coli* according to standard techniques known in the fields of molecular biology, bioengineering, and genetic engineering.

A broad-host range plasmid encoding pdxJ can be introduced into *S. meliloti* IFO 14782 by
15 tri-parental mating in the following manner. *S. meliloti* as a recipient strain, *E. coli* harboring helper plasmid as a helper strain, and *E. coli* harboring donor plasmid as a donor strain are cultivated separately and mixed together. After mixed cultivation on plate, *S. meliloti* receiving a recombinant plasmid can be selected on agar plate containing appropriate antibiotics. The plasmids of colonies grown on the plates are examined by endonuclease di-
20 gestion.

(2) Preparation of Mutant Endowed with Amino Acid Requirement Induced by Mutagenesis with N-methyl-N'-nitro-N-nitrosoguanidine (referred to as NTG hereinafter)

Pyridoxol in *S. meliloti* IFO 14782 is known to be synthesized by ring closure of two precursors, 1-deoxy-D-xylulose and 4-hydroxy-L-threonine [Tazoe et al., J. Biol. Chem.
25 275:11300-11305 (2000)]. In general, accumulation of amino acids synthesized by a branched pathway is reported to be greatly enhanced by induction of amino acid requirement. Thus it is conceivable to isolate amino acid requiring mutants to get higher vitamin B₆ producer. *S. meliloti* IFO 14782/pVKP601 prepared in (1) [B] is subjected to NTG mutagenesis to produce mutants producing pyridoxol more in the culture broth by induc-
30 tion of amino acid requiring mutants. Cells of the strain are treated with NTG. After treatment, a restorative cultivation is carried out and the resulting culture is plated out on agar medium. To isolate mutants requiring amino acid, the growth of colonies is tested on agar of inorganic nitrogen salt medium containing vitamins and nucleic acids. From the test, colonies requiring amino acid can be selected, and a vitamin B₆ high producer may be

selected by testing productivity of vitamin B₆ in the fermentation. The strain PY-C341-K1 is one of the objective mutants in this present invention.

(3) Preparation of Mutants Endowed with Glycine Resistance Induced by Mutagenesis with NTG

- 5 Biosynthesis of vitamin B₆ in *S. meliloti* IFO 14782 is well-known as described in (2). Pyridoxol is synthesized from a sugar and an amino acid precursors, and the latter precursor is from glycolaldehyde and glycine. Glycine is not only one of vitamin B₆ precursors but also a strong inhibitor to the growth of the strain. Accordingly, induction of glycine resistant mutant leads to enhancement of production of vitamin B₆. To isolate
10 glycine resistant mutants, minimal inhibitory concentration of glycine against the strain should be examined on an appropriate medium because the inhibition strength was different in tested medium. Thus, strain PY-C341-K1 obtained in (2) is subjected to NTG mutagenesis to produce glycine resistant mutants. Cells of the strain are treated with NTG in a similar manner as described in (2). After treatment, a restorative cultivation is carried
15 out and the resulting culture is plated out on an agar medium. To isolate glycine resistant mutants, the cell suspension is spread onto plates of agar medium containing appropriate concentration of glycine. After incubation, colonies resistant to glycine may be selected on agar medium containing glycine.

- The microorganisms obtained in the present invention are incubated in a medium
20 containing an assimilable carbon source, a digestible nitrogen source, an inorganic salt, and other nutrients necessary for their growth. As a carbon source, for example, glucose, fructose, lactose, maltose, galactose, sucrose, starch, dextrin, or glycerol may be employed. As a nitrogen source, for example, peptone, corn steep liquor, soybean powder, yeast extract, meat extract, ammonium chloride, ammonium sulfate, ammonium nitrate, urea,
25 or their mixture thereof may be employed. Further, for trace elements, sulfates, hydrochlorides, or phosphates of calcium, magnesium, zinc, manganese, cobalt, and iron may be employed. And, if necessary, conventional nutrient factors, a trapping agent of phosphate ion, or an antifoaming agent, such as magnesium carbonate, aluminum oxide, allophane, animal oil, vegetable oil, or mineral oil can also be added supplementary in a
30 fermentation medium.

The pH of the culture medium may be about 5.0 to 9.0, preferably 6.5 to 7.5. The cultivation temperature may be about 10°C to 40°C, preferably 25°C to 35°C. The cultivation time may be about 1 day to 15 days, preferably 2 days to 9 days.

In the cultivation, aeration and agitation usually give favorable results.

After the cultivation, vitamin B₆ produced also may be separated from the culture broth and purified. For this purpose a process generally used for extracting a certain product from the culture broth may be applied by utilizing various properties of vitamin B₆. Thus, 5 for example, the cells are removed from the culture broth, the desired substance in the filtrate is absorbed on active carbon, then eluted and purified further with an ion exchange resin. Alternatively, the culture filtrate is applied directly to an ion exchange resin and, after the elution, the desired product is recrystallized from mixture of alcohol and water.

The microorganisms used in the present invention include all the mutant strains of genus 10 *Sinorhizobium* capable of producing vitamin B₆ having a recombinant plasmid with *pdxJ* gene that acquired phenotypic property of histidine requirement, or glycine resistance, or combination of their phenotypic properties thereof. Among the strains of genus *Sinorhizobium*, a particularly preferred strain is *S. meliloti* PY-EGC1, which was deposited on September 17, 2002 under deposit number DSM15209 at the DSMZ (Deutsche 15 Sammlung von Mikroorganismen und Zellkulturen GmbH) in Göttingen (Germany) under the Budapest Treaty.

The present invention will be explained more in detail by referring to the following examples; however, it should be understood that the present invention is not limited to those particular examples.

20 In the Examples, the amount of vitamin B₆ produced in culture broth can be assayed by the turbidity method with *Saccharomyces carlsbergensis* ATCC 9080 [Osborne and Voegt, The Analysis of Nutrients in Foods, Academic Press, London, 224-227 (1978)], and vitamin B₆ derivatives such as pyridoxol, pyridoxal, and pyridoxamine in a fermentation broth can be separately quantified by high pressure liquid chromatography (referred to as 25 HPLC hereinafter).

Example 1: Preparation of *S. meliloti* IFO 14782/pVK601

(1) Cloning of *pdxJ* of *S. meliloti* IFO 14782

To amplify *pdxJ* of *S. meliloti* IFO 14782 using PCR method, the following two primers were synthesized according to the DNA sequence of *pdxJ* (2249854-2250606, comple- 30 ment) in the genome database of *S. meliloti* strain 1021 (Accession No. NC_003047): primer A (SEQ ID NO:1) with restriction enzyme *NdeI* recognition sequence including start codon of *pdxJ* and primer B (SEQ ID NO:2) with *PstI* site just after stop codon of *pdxJ*. Chromosomal DNA was extracted from the cells grown in a medium (referred to as

LBMC hereinafter) composed of 1 % Bacto Tryptone (Becton Dickinson Microbiology systems, MD, USA), 0.5 % Bacto Yeast extract (Becton Dickinson Microbiology systems, MD, USA), 0.5 % NaCl, 0.061 % $MgSO_4 \cdot 7H_2O$, and 0.036 % $CaCl_2 \cdot 2H_2O$ with QIAGEN genomic-tips (QIAGEN GmbH, Germany).

5 PCR was performed using advantage-HF PCR kit (CLONTECH Laboratories, Inc. CA, USA). 100 μ l of reaction mixture contained 10 ng of chromosomal DNA of *S. meliloti* IFO 14782, 50 pmol of the two primers, 10 μ l of 10 \times HF dNTP mix, 10 μ l of appended 10 \times HF PCR reaction buffer, and 2 μ l of 50 \times advantage-HF polymerase mix. The reaction conditions were as follows; holding at 94°C for 3 min., 4 cycles of 30 sec at 98°C, 1 min at 10 53°C, 1 min at 72°C, 20 cycles of 30 sec at 98°C, 1 min 68°C, and holding at 72°C for 10 min. 10 μ l of reaction mixture was subjected to agarose gel on 1 % (w/v) gels, and a DNA band of 770 bp was recovered from the gel with QIAEXII (QIAGEN GmbH, Germany). The fragment was ligated to pUC18, which was digested with SmaI and dephosphorylated with alkaline phosphatase, by SureClone ligation kit (Amersham Biosciences Corp., NJ, 15 U.S.A.).

The thus obtained ligation mixture was transformed into *E. coli* JM109 competent cells (Takara Bio Inc., Shiga, Japan) and plated on plates of a medium composed of 1 % Bacto Tryptone, 0.5 % Bacto Yeast extract, and 0.5 % NaCl (referred to as LB hereinafter) containing 100 μ g/ml of ampicillin (referred to as Amp). Plasmids of colonies grown on the 20 plates were prepared with Automatic DNA Isolation System PI-50 (Kurabo Industry Ltd., Japan). By analysis of the plasmid with restriction enzyme, a recombinant plasmid pSHT56, wherein pdxJ was the same direction as *lacZ* gene on pUC18, was obtained. pSHT56 was prepared from *E. coli* JM109 harboring pSHT56 with QIAGEN plasmid Midi kit (QIAGEN GmbH, Germany). The DNA sequence of pdxJ in the plasmid was 25 ascertained with an ALF DNA sequencer (Amersham Biosciences Corp., NJ, U.S.A.) and it was identical with that of a genome database of *S. meliloti* strain 1021.

As a vector for expression of pdxJ in *E. coli*, pUC 18 was remodeled into pUC-trc2, which has *trc* promoter region of pTrc99A (Amersham Biosciences Corp., NJ, USA) followed by NdeI recognition sequence. pUC-trc2 was prepared from *E. coli* JM109 harboring pUC-trc2 with QIAGEN plasmid Midi kit. To give an expression plasmid for pdxJ in *E. coli*, 30 pUC-trc2 was digested with NdeI, and a 2.5-kb fragment was recovered from agarose gel and dephosphorylated with alkaline phosphatase (Takara Bio Inc., Shiga, Japan). On the other hand, pSHT56 was cleaved with NdeI, subjected to agarose gel and resulting 1-kb fragment was recovered from the gel with QIAEXII. The recovered 1-kb fragment 35 was ligated to prescribed 2.5-kb fragment of pUC-trc2 with ligation kit (Takara Bio Inc.,

Shiga, Japan). *E. coli* JM109 was transformed with thus-obtained ligation mixture and plated on LB plates containing 100 µg/ml of Amp. Plasmid of a colony grown on the plate was prepared with Automatic DNA Isolation System PI-50. By analysis of the plasmid with restriction enzyme, a recombinant plasmid pSHT57, wherein *pdxJ* was the same direction as *trc* promoter, was obtained (Figure 1). pSHT57 was prepared from *E. coli* JM109 harboring pSHT57 with QIAGEN plasmid Midi kit.

(2) Construction of expression plasmid for *pdxJ* in *S. meliloti* IFO 14782

To construct an expression vector in *S. meliloti* IFO 14782, pVK100 was used, which is reported to be a broad host range vector, IncP-1 type, and replicable in *S. meliloti*. pVK100 was prepared from *E. coli* HB101/pVK100 with QIAGEN plasmid midi kit, and digested with HindIII, blunt-ended by blunting kit (Takara Bio Inc., Shiga, Japan) and dephosphorylated with alkaline phosphatase. pSHT57 was digested with BamHI and KpnI. Resulting 875-bp fragment, which contained *trc* promoter and *pdxJ*, was recovered from agarose gel, blunt-ended by blunting kit, and ligated to prescribed pVK100 with ligation kit. *E. coli* HB101 competent cells (Takara Bio Inc., Shiga, Japan) was transformed with the obtained ligation mixture and plated on LB plates containing 10 µg/ml of tetracycline (referred to as Tc hereinafter). Plasmids of colonies grown on the plates were prepared with Automatic DNA Isolation System PI-50. By analysis of the plasmid with restriction enzyme, a recombinant plasmid, pVK601, wherein *trc* promoter and *pdxJ* were the opposite direction against kanamycin resistant gene on, was obtained (Figure 1).

(3) Complementing *E. coli* AT3208, *pdxJ*⁻ mutant

pSHT57 was transformed into *pdxJ*⁻ mutant, *E. coli* AT3208 (purchased from *E. coli* Genetic Stock Center, Yale Univ., U.S.A.). All Amp resistant transformants grew on vitamin B₆-free EMM plate composed of 10 g of glucose, 8 g of vitamin-free casamino acid (Becton Dickinson Microbiology systems, MD, U.S.A.), 2.5 mg of MnSO₄·5H₂O, 125 mg of MgSO₄·7H₂O, 125 mg of CaCl₂·2H₂O, 425 mg of KCl, 250 µg FeCl₃·6H₂O, 250 µg of thiamin hydrochloride, 8 µg of biotin, 15 g of Bacto agar (Becton Dickinson Microbiology systems, MD, U.S.A.) per liter (pH 6.8) whereas *pdxJ*⁻ mutant, *E. coli* AT3208, did not grow on the plate. This indicated that cloned *pdxJ* worked as PdxJ.

(4) Introduction of pVK601 into *S. meliloti* IFO 14782

pVK601 was introduced into *S. meliloti* IFO14782 by tri-parental mating as described below. *S. meliloti* IFO14782 as a recipient strain was inoculated in 5 ml of liquid LBMC medium and incubated with shaking at 30°C at 140 rpm for 16 hours. 400 µl of the

culture were transferred into the fresh same medium and incubated further for 6 hours. *E. coli* HB101 harboring pRK2013 (Km^r ; IncP tra^+ , ColEI ori)(ATCC 37159) as a helper strain was inoculated in 5 ml of liquid LB medium containing 50 μ g/ml of kanamycin and incubated with shaking at 37°C at 140 rpm for 16 hours. 100 μ l of the culture were transferred into the fresh same medium and incubated further for 6 hours. *E. coli* HB101 harboring pVK601 was inoculated in 5 ml of liquid LB medium containing 10 μ g/ml of Tc and incubated with shaking at 37°C at 140 rpm for 16 hours. 100 μ l of the culture were transferred into the fresh same medium and incubated further for 6 hours. Each strain was harvested and cells were mixed at 1:1:4 (v/v/v) ratio. The mixture was put on a nitrocellulose filter placed on LBMC agar plate. After this plate was incubated for 20 hours at 30°C, cells on the filter were scratched and suspended in sterilized 0.85 % NaCl solution. The suspension was diluted appropriately and spread on LBMC plates containing 20 μ g/ml nalidixic acid (to select for *S. meliloti* IFO14782) and 10 μ g/ml Tc (to select for pVK601). After incubation of these plates at 30°C for 5 days, colonies grown on the plates were picked up and cultured for plasmid extraction by QIAGEN plasmid mini kit (QIAGEN GmbH, Germany). Thus-obtained plasmid DNA with treatment of endonuclease showed an identical pattern to pVK601 on agarose gel. From this result, one colony in the tested colonies was selected as *S. meliloti* IFO 14782/pVK601.

Example 2

A loopful of *S. meliloti* IFO14782 / pVK601 and the parent, *S. meliloti* IFO14782, grown on a LBMC agar plate at 30°C for 48 hours was inoculated to tubes containing 8 ml of a seed medium (referred to as SM hereinafter) composed of 1% glucose, 1 % corn steep liquor (Oji Cornstarch Co., Ltd., Tokyo, Japan), 0.2 % Bacto yeast extract, 0.05 % $MgSO_4 \cdot 7H_2O$, 0.001 % $MnSO_4 \cdot 5H_2O$, and 0.001 % $FeSO_4 \cdot 7H_2O$ (pH 7.0) and then the tubes were shaken on a reciprocal shaker (275 rpm) at 30°C. After shaking for 19 hours, each 4 ml of the cultures was transferred to a 500 ml flask with two baffles containing 200 ml of a production medium (referred to as PM hereinafter) composed of 6 % glucose, 3 % corn steep liquor, 0.8 % Bacto yeast extract, 0.35 % NH_4Cl , 0.05 % $MgSO_4 \cdot 7H_2O$, 0.025 % $MnSO_4 \cdot 5H_2O$, 1 % Allophosite (Shinagawa Chemicals Co., Ltd., Tokyo, Japan) and 0.025 % Actocol (pH 6.8) and shaken on a rotary shaker (180 rpm) at 30°C. After cultivation for 7 days, contents of vitamin B₆ in the supernatant of each culture broth were quantified by high pressure liquid chromatography (referred to as HPLC hereinafter) and produced vitamin B₆ was calculated by the internal standard method with 4'-deoxypyridoxol as

described below. To prepare the samples for HPLC, 100 μ l of the solution containing 100 mg/l of 4'-deoxypyridoxol as internal substance was added to 400 μ l of the standard solutions of pyridoxol and the supernatant from the culture broth, and then the mixture was put on the following column. The analytical conditions were as follows: column,

5 Capcell pak C18 SG120 (4.6 \times 250 mm) (Shiseido Co., Ltd., Tokyo, Japan); mobile phase, 0.1 M sodium perchlorate, 0.1 M potassium phosphate, and 2 % acetonitrile (pH 3.5); column temperature, 25–26°C; flow rate, 1.0 ml/min; and detector, ultraviolet (UV) (at 292 nm). As a result, *S. meliloti* IFO14782 /pVK601 produced 119 mg of pyridoxol per liter and was about 1.34 times higher than the parent, strain IFO 14782.

10 Example 3

S. meliloti IFO14782 /pVK601 obtained in Example 1 was cultured in a flask containing LBMCG containing 5 μ g/ml of Tc for 17 hours at 30 °C, and the cell suspension of the strain was prepared. A tube containing 5 ml of the reaction mixture composed of 150 μ g/ml of NTG and 1.6×10^9 cells per ml in 50 mM Tris-HCl buffer (pH 8.0) was incubated

15 with a reciprocal shaking (275 rpm) for 30 min at 30°C. The cells were washed twice with sterile saline and suspended in saline. 100 μ l of the cell suspension was spread onto agar plates containing LBMCG containing 5 μ g/ml of Tc, and then the plates were incubated for 2 days at 30°C. The cells grown on the plates were recovered in 7 ml of sterile saline, and the cell suspensions were serially diluted 10^{-1} – 10^{-7} in sterile saline and spread onto the

20 same agar plates. To isolate amino acid requiring mutants, the growth of 1,136 colonies grown on LBMCG containing 5 μ g/ml of Tc were tested on agar of a minimum medium (referred to as MM hereinafter) composed of 1 % glucose, 0.22 % NH_4Cl , 0.06 % K_2HPO_4 , 0.06 % KH_2PO_4 , 0.04 % $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$, 0.02 % $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$, 0.04 % NaCl , and the following metal salts, vitamins, and nucleic acids (per liter): 12 mg of $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$, 4 mg of

25 $\text{MnSO}_4 \cdot 5\text{H}_2\text{O}$, 0.5 mg of H_3BO_3 , 0.4 mg of Na_2MoO_4 , 0.32 mg of $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$, 0.04 mg of $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$, 0.002 mg of $\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$, 4 mg of calcium pantothenate, 3 mg of thiamin-HCl, 1.25 mg of riboflavine, 0.04 mg of biotin, 10 mg of hypoxanthine, 10 mg of guanine sulfate, 10 mg of thymine, and 10 mg of uracil containing 5 μ g/ml of Tc. After incubation for 4 days at 30°C, 37 colonies indicating a poor or no growth were picked up to

30 LBMCG agar containing 5 μ g/ml of Tc, and the productivity of vitamin B₆ was examined by flask fermentation. One loopful cells of 37 colonies and *S. meliloti* IFO14782 /pVK601 (parent) were inoculated to tubes containing 8 ml of SM medium, and then the tubes were shaken on a reciprocal shaker (275 rpm) at 30°C. After shaking for 19 hours, each 4 ml of

culture broth was transferred to 500-ml flasks with two baffles containing 200 ml of PM medium and shaken on a rotary shaker (180 rpm) at 30°C. After cultivation for 7 days, contents of vitamin B₆ in the supernatant of each culture broth were quantified by HPLC. As a result, *S. meliloti* PY-C341K1 produced 171 mg of pyridoxol per liter and was 1.44 times higher than the parent, strain IFO14782 /pVK601.

The amino acid requirement of strain PY-C341K1 together with the parent, strain IFO14782/pVK601, was examined by culturing for 2 days at 30°C in tubes containing 8 ml of MM supplemented with various kinds of amino acids. From a result, strain PY-C341K1 grew as well as the parent, IFO14782 /pVK601, in MM medium supplemented with 42 µg/ml of histidine.

Example 4

In a similar manner as described in Example 3, *S. meliloti* PY-C341K1 was cultured in a flask containing LBMCG containing 10 µg/ml of Tc for 16 hours at 30°C, and the cell suspension of the strain was prepared. A tube containing 5 ml of the reaction mixtures composed of 0, 30, and 50 µg/ml of NTG and 1.6×10^9 cells per ml in 50 mM Tris-HCl buffer (pH 8.0) was incubated with a reciprocal shaking (275 rpm) for 30 min at 30°C. The cells of each reaction mixture were washed twice with sterile saline and suspended in saline. 100 µl of the cell suspension was spread onto agar plates containing LBMCG containing 10 µg/ml of Tc, and then the plates were incubated for 2-3 days at 30°C. The cells grown on the plates were recovered by suspending in sterile saline. After centrifugation of the suspension, the cell suspension was diluted to give a turbidity of OD₆₀₀ = 1.6, and finally to 10⁻⁵. Each 100 µl of the diluents was spread onto five agar plates containing LBMCG containing 10 µg/ml of Tc and 0, 0.125, 0.15, or 0.175 % glycine because 0.15 % glycine completely inhibited the growth of *S. meliloti* PY-C341K1 on LBMCG plate, and then the plates were incubated for 4 days at 30°C. Ten colonies treated with 50 µg/ml of NTG grown on plates LBMCG containing 10 µg/ml of Tc and 0.175 % glycine were picked up on LBMCG agar containing 10 µg/ml of Tc. After incubation for 2 days at 30°C, the productivity of vitamin B₆ in ten colonies together with the parent strain (*S. meliloti* PY-C341K1) was examined by flask fermentation. One loopful cells was inoculated to tubes containing 8 ml of SM medium, and then the tubes were shaken on a reciprocal shaker (275 rpm) at 30°C. After shaking for 19 hours, each 4 ml of culture broth was transferred to a 500-ml flask with two baffles containing 200 ml of PM medium modified to 0.175 % NH₄Cl, and shaken on a rotary shaker (180 rpm) at 30°C. After shaking for 4 days, sterile

solution of urea was added to the each flask at 0.125 %, and the shaking were further continued for 3 days. The contents of vitamin B₆ in the supernatant of 7-day culture broth were quantified by HPLC method as described in Example 3. As a result, *S. meliloti* PY-EGC1 produced 362 mg of pyridoxol per liter and was about 2.11 times higher than strain PY-341K1 (the parent).

Example 5

Vitamin B₆ was recovered from the culture broth of *S. meliloti* PY-EGC1 prepared in the same cultural conditions as described in Example 5. Pyridoxol at each purification step and the concentration was followed by HPLC. One liter of the 168 hour-culture broth containing 344 mg/L of PN was centrifuged at 7,500 rpm for 10 min. The pH of the resultant supernatant was adjusted to 3.1 with 1N hydrochloric acid, and then the supernatant was applied to a column (5.5 × 15 cm) packed with 350 ml of Amberlite CG 120 (H⁺ form, 100–200 mesh, Rohm and Haas Company, Philadelphia, Pennsylvania, USA). The column was washed with 500 ml of deionized water and then eluted with 5% ammonium hydroxide. The vitamin B₆ fractions were concentrated under reduced pressure. The residue thus obtained was dissolved in 10 ml of deionized water, and the solution was charged on a column (5.5 × 16 cm) packed with 380 ml of Dowex 1 × 4 (OH⁻ form, 200–400 mesh, Dow Chemical Co., Ltd., Midland, Michigan, U.S.A.), and then washed with 500 ml of deionized water. The column was then eluted with 0.1 N HCl. The fractions containing pyridoxol was concentrated to small volume under reduced pressure. After the solid residue was dissolved in small amount of hot ethanol, the solution was kept standing at 4°C overnight. The resultant precipitates were collected by filtration and dried in vacuo to obtain 282 mg of crude crystals. It was recrystallized from ethanol to obtain 217 mg of white crystals having a melting point of 160°C. The infrared absorption, UV absorption, and NMR spectrum of the product of the product coincided with those of authentic pyridoxol.

Table 1 summarizes the vitamin B₆ productivities of *S. meliloti* IFO 14782 (DSM No. 10226), *S. meliloti* IFO 14782/pVK601, and their mutants so far obtained.

Table 1: Pyridoxol Productivities of *S. meliloti* IFO 14782 (DSM No. 10226), *S. meliloti* IFO 14782/pVKP601, and their Mutants

Example	Microorganism	Phenotypic properties	Pyridoxol (mg/L)	Magnitude of increasing
	<i>S. meliloti</i> IFO 14782 (DSM 10226)		89	1.0
2	<i>S. meliloti</i> IFO 14782/pVKP601	amplification of <i>pdxJ</i>	119	1.34
3	<i>S. meliloti</i> PY-C341K1	histidine requirement	171	1.92
4	<i>S. meliloti</i> PY-EGC1	glycine resistance	362	4.07

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Claims

1. A mutant of a recombinant microorganism of the genus *Sinorhizobium* capable of producing vitamin B₆ having a recombinant plasmid with *pdxJ* gene that acquired histidine requirement or glycine resistance, or its combination thereof.
- 5 2. The mutant according to claim 1, wherein said mutant possesses resistance to glycine of 1.1-fold higher than the concentration of minimal inhibitory concentration of the recombinant microorganism.
3. The recombinant microorganism according to claim 1, wherein said recombinant plasmid comprises vector plasmid pVK100.
- 10 4. The recombinant microorganism according to claim 1, wherein pyridoxol 5'-phosphate synthase gene is derived from microorganism of the genus *Sinorhizobium* which is capable of producing vitamin B₆.
5. The recombinant microorganism according to claim 1 which is *Sinorhizobium meliloti* PY-EGC1.
- 15 6. A process for producing vitamin B₆ which comprising cultivating the microorganism according to claim 1 in a cultivation medium at a pH value of about 5.0 to 9.0, at a temperature of 10°C to 40°C, and for 1 day to 15 days under aerobic conditions, isolating vitamin B₆ from the cultivation medium.
- 20 7. The process according to claim 6, wherein the microorganism is *Sinorhizobium meliloti* PY-EGC1.

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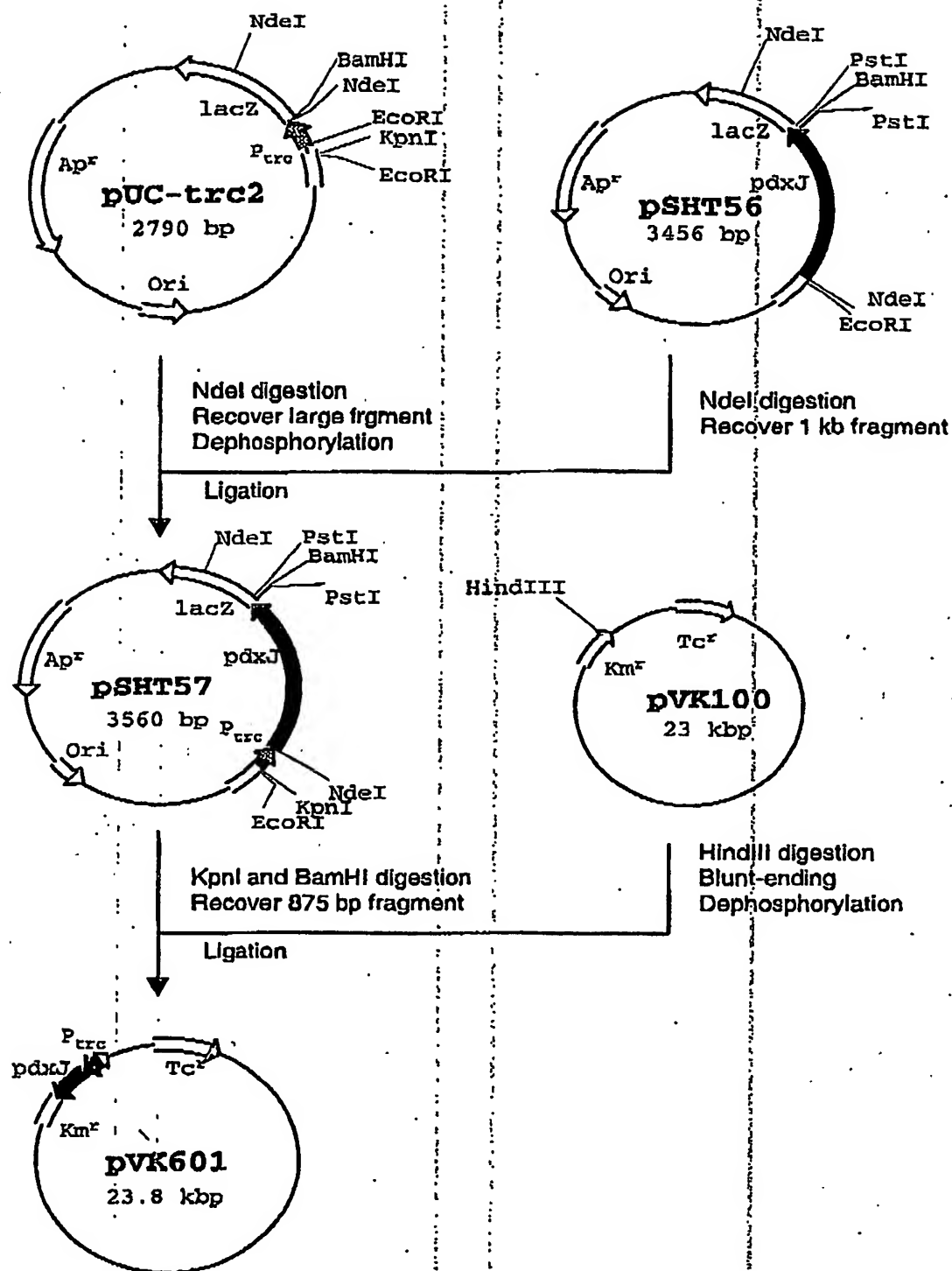


Fig. 1 The construction of a recombinant plasmid pVKP601

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